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Novel Synthesis of Nectrisine and 4-epi-Nectrisine

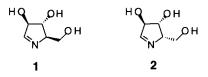
Yong Jip Kim and Takeshi Kitahara*

Department of Applied Biological Chemistry, Graduate School of Agricultural and Life Sciences, The University of Tokyo, 1-1-1 Yayoi, Bunkyo-ku, Tokyo 113, Japan.

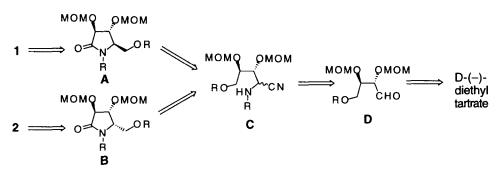
Abstract: Nectrisine 1, a potent glycosidase inhibitor, and 4-epi-Nectrisine 2 were synthesized from D(-)- diethyl tartrate through the corresponding lactams. N-acyl protection was crucial to effect the reduction of the corresponding lactams to amino alcohols © 1997 Elsevier Science Ltd.

Much attention has been paid to polyhydroxylated pyrrolidines since they possess remarkable biological activities as inhibitors of glycosidases.^{1,2} One such substance, nectrisine 1, a fungal metabolite isolated from *Nectria lucida*, has attracted significant interest.³ Syntheses of 1 have been published, mostly starting from sugars as chiral precursors.⁴

Herein we wish to describe the novel synthesis of nectrisine 1 and 4-*epi*-nectrisine 2 from D-(–)-diethyl tartrate *via* lactam intermediates 9a, 9b respectively. Although 4-*epi*-nectrisine 2 has been reported to exist as a complex equilibrium mixture with dimerized and hydrated forms, its biological activities are not known.⁵



Our synthetic strategy for a general approach to both diastereomers of nectrisine 1 and 4-epi-nectrisine 2 is illustrated in Scheme 1. 1 and 2 can be envisioned as being derived from the lactam precursors A and B by reduction of the carbonylfunctions, followed by hydrolytic deprotection of the protecting groups. Diastereomeric pyrrolidinones A and B can be generated from aminonitrile C by oxidative lactam formation and



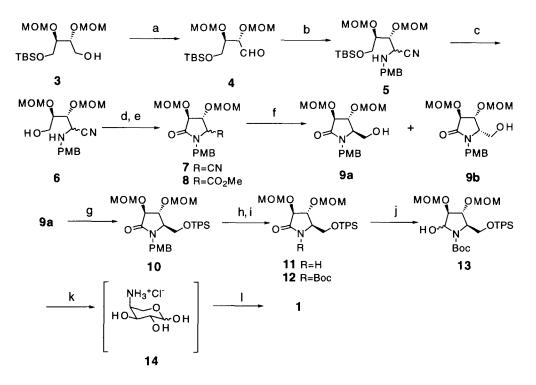
Scheme 1

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reduction of the nitrile. The diastereomeric mixture of amino nitriles C can be easily prepared by a modified Strecker reaction⁶ with aldehyde **D** provided from the starting material D-(-)-diethyl tartrate.

Our synthesis began with Swern oxidation of the known alcohol $3,^7$ which was obtained from D-(-)diethyl tartrate in good yield (3steps, >90%) and is depicted in Scheme 2. Reaction of this aldehyde 4 with 2.4 eq. of *p*-methoxybenzylamine and 1.2 eq. of diethyl phosphorocyanidate (DEPC)⁶ in THF gave aminonitrile 5 (2steps, 96%), as an inseparable diastereomeric mixture which was subsequently deprotected with tetra-*n*butylammonium fluoride (TBAF) in THF to the corresponding amino alcohol 6 in 87% yield. The amino alcohol 6 was oxidized by reaction with tetra-*n*-propylammonium perruthenate (TPAP)⁸ and *N*methylmorpholine-N-oxide (NMO) in dichloromethane at room temperature for 40 min to give the lactam 7.

Without purification, the lactam was treated with 3 eq. of sodium methoxide in methanol at room temperature to give methyl ester 8 (2steps, 71%), which was reduced with $LiBH_4$ (lithium borohydride) in THF to form an alcohol and produced a chromatographically separable mixture of two diastereomers in 87% yield, *trans*-lactam 9a and *cis*-lactam 9b, in a ratio of 56:44. Protection of the primary alcohol of *trans*-lactam 9a with *t*-butyldiphenylsilylchloride (TPSCI) and Et₃N gave the silyl ether 10 in 96% yield.

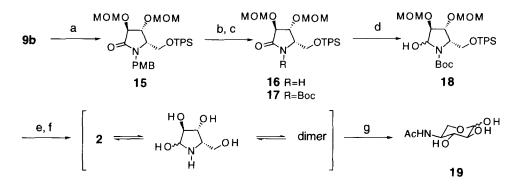


a) Swern oxidation; b) p-(CH₃O)C₆H₄CH₂NH₂, (EtO)₂P(O)CN, THF; 2 steps, 96%; c) TBAF, THF; 87%; d) TPAP, NMO, MS4A, CH₂Cl₂; e) NaOMe, MeOH, 0°C \rightarrow r.t. then 1N HCl ; 2 steps, 71%; f) LiBH₄, THF, 0°C \rightarrow r.t.; 87%; g) imidazole, TPSCl, DMF; 96%; h) (NH₄)₂Ce(NO₃)₆, CH₃CN-H₂O (9:1), 0°C; 84%; i) Et₃N, (Boc)₂O, DMAP, CH₂Cl₂; quant.; j) LiEt₃BH, THF, -78°C; 93%; k) 6N HCl, THF, 50°C, 2h; >80%, l) Dowex 1-X2 (OH⁻), 90%

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Scheme 2
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The key step in this synthesis is the reduction of lactam to amino alcohol. Accordingly, we examined the reduction of **10** with various reducing reagents, but our attempts were unsuccessful. Faced with this problem, we decided to replace the *N*-protecting group, PMB, with the more electron-withdrawing and easily removable Boc group. Removal of the PMB protecting group in **10** by ceric ammonium nitrate (CAN)⁹ in CH₃CN-H₂O (9:1) at 0°C provided lactam **11** in 84% yield, which was treated with di-*t*-Butyl dicarbonate (Boc₂O) and Et₃N in CH₂Cl₂ to give an imide **12** in quantitative yield. Thus, reduction of the imide **12** with LiEt₃BH (Super Hydride[®])¹⁰ in THF at -78°C cleanly afforded amino alcohol **13** in 93% yield. The final task was the removal of the protecting groups. This was accomplished by treatment of 6N HCl in THF at 50°C for 2h to give the amino sugar precursor **14**¹¹ (>80% yield), followed by ion exchange chromatography (Dowex resin, OH=form) which afforded nectrisine **1** in 90% yield. Comparison of the specific optical rotation, ¹H and ¹³C NMR data of our synthetic nectrisine **1**¹² with those in the literature^{3,4b} completely confirmed the identity of nectrisine.

The 4-*epi*-nectrisine **2** was then synthesized from **9b**, following the set of reactions previously described for the nectrisine **1**. Thus, as depicted in Scheme 3, protection of the primary hydroxyl function with TPSCI and Et₃N gave the silyl ether **15** in 92% yield. Removal of the protecting PMB group by CAN in CH₃CN-H₂O (9:1) at 0°C gave lactam **16** in 82% yield, which was then treated with Boc₂O and Et₃N in CH₂Cl₂ to give an imide **17**(quant. yield). The imide was reduced to amino alcohol **18** by Super Hydride[®] in THF at -78°C in 95% yield. Finally, deprotection of **18** with 6N HCl in THF at 50°C for 2h yielded a mixture of products (>80% yield). 4-*epi*-Nectrisine **2** was reported to exist as an equilibrium mixture of several forms.⁵ For identification, we decided to acetylate the amino group. Treatment of the mixture of products with Ac₂O in H₂O gave **19**¹³ (quant. yield) which was identical in all respects with that in the literature.^{5a)}



a) imidazole, TPSCl, DMF; 92%; b) $(NH_4)_2Ce(NO_3)_6$, $CH_3CN-H_2O(9:1)$, 0°C; 82%; c) Et₃N, $(Boc)_2O$, DMAP, CH_2Cl_2 ; quant.; d) LiEt₃BH, THF, -78°C; 95%; e) 6N HCl, THF, 50°C, 2h; f) Dowex 1-X2 (OH⁻); >80%, g) Ac₂O, H₂O; Dowex 50W-X2 (H⁺); quant.

Scheme 3

In summary, a new route to 1 and 2 via lactam intermediates 9a, 9b has been developed. These pathways produce various synthetic intermediates and analogs which may be helpful in evaluating structureactivity relationships of this glucosidase inhibitor. A thorough biochemical evaluation of 2 and related derivatives is in progress and will be reported elsewhere.

Acknowledgments

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- 12. Physical data for 1: $[\alpha]_{D}^{20} = +19.6^{\circ}(c=0.5, H_{2}O)$, lit : $[\alpha]_{D} = +21.8^{\circ}(c=0.6, H_{2}O)$, ^{4b} lit : $[\alpha]_{D}^{23} = +22.0^{\circ}(c=0.55, H_{2}O)$, ³ IR (KBr): 3300, 2900, 1640, 1560, 1400, 1040, 850 cm⁻¹, ¹H NMR (300 MHz, D₂O) δ 7.67 (1H, bs), 3.09-4.08 (5H, m), ¹³C NMR (300 MHz, D₂O) δ 170.7, 83.6, 78.5, 77.0, 61.4.
- 13. Physical data for 19: [α]_D²⁰ = -47.7° (c=0.9, H₂O), lit : [α]_D²⁴ = -53° -49° (c=3.3, H₂O),^{5a} IR (Nujol): 3330, 2920, 2850, 1710, 1640, 1560, 1460, 1380, 1070 cm⁻¹. ¹H NMR (500 MHz, D₂O) δ 5.24 (α-anomer, 0.4H, d, *J*=3.5Hz), 4.57 (β-anomer, 0.6H, d, *J*=7.5Hz), 3.89 (2H, m), 3.67 (2H, m), 3.31 (1H, m), 2.02 (3H, s), ¹³C NMR (500 MHz, D₂O) δ 175.3, 97.3, 93.1, 75.4, 74.1, 72.7, 70.9, 64.2, 60.0, 51.8(2C), 22.7.

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